

# Data analysis for solution scattering of biomacromolecular complexes

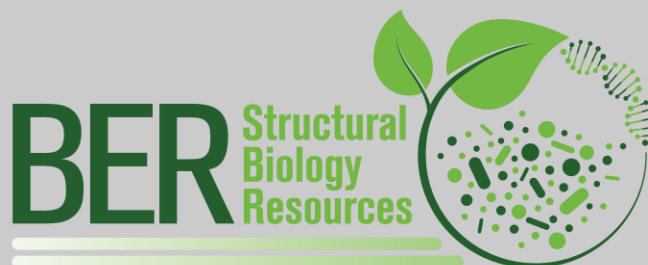
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Neutron Scattering Division

Oak Ridge National Laboratory

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**ENERGY**

# Topics to be covered

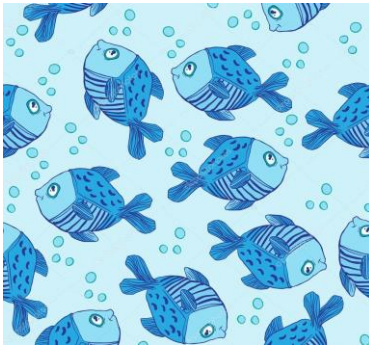
- Data analysis for solution scattering
  1. Guinier Analysis
  2.  $P(r)$  calculation
  3. Kratky Plot
  4. Mw Estimation
  5. Fitting experimental data with PDB structures
  6. Software for modeling SANS data

# Resources

- Data analysis software ATSAS 3.1.1 - <https://www.embl-hamburg.de/biosaxs/software.html>
  - <https://www.embl-hamburg.de/biosaxs/atsas-online/>
  - CRYSON
  - GNON
  - SAXSREF/CV
  - OLIGOMER
  - DAMMIN/DAMMIF
  - GASBOR
  - MONSA
  - EOM
  - SUPCOMP
- BioXTAS RAW - <https://bioxtas-raw.readthedocs.io/en/latest/index.html>
  - GNON
  - DAMMIN/DAMMIF
  - DENSS
- ScÅtter - <https://bl1231.als.lbl.gov/scatter/>
- Protein data bank - <https://www.rcsb.org/>
- AlphaFold - <https://colab.research.google.com/github/deepmind/alphafold/blob/main/notebooks/AlphaFold.ipynb>
- Pymol – <https://www.pymol.org/2/>
- Chimera - <https://www.cgl.ucsf.edu/chimera/>
- FoxSAXS - <https://modbase.compbio.ucsf.edu/foxs/>

# Monodispersed samples for solution scattering

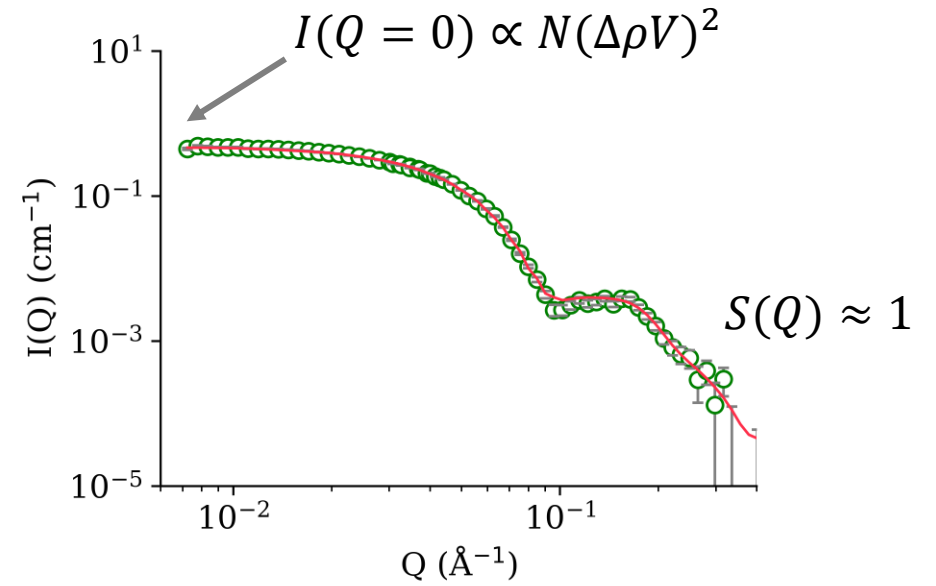
Homogeneous particles



$$I(q) = S(q) \sum_i^n [(\Delta\rho_i V_i)^2 P_i(q)].$$

1. Pair distance distribution - Dmax
2. Overall size – Rg
3. Conformation – Globular vs extended
4. Oligomerization state
5. Molecular mass – MW
6. Comparison with PDB structures (AlphaFold)
7. Molecular envelope

Scattering 1D profile of monodispersed sample



## Experiment details:

- Protiated protein in 100% D<sub>2</sub>O Buffer
- Buffer exchange to 100% D<sub>2</sub>O applying concentrations/dilution cycles
- Concentration used – 2 mg/mL (41.7 KDa)
- Data collected for 2 h (good S/N)
- Volume used 320 uL
- Oligomer: Hexamer

# Background subtraction

First step - Empty cell subtraction

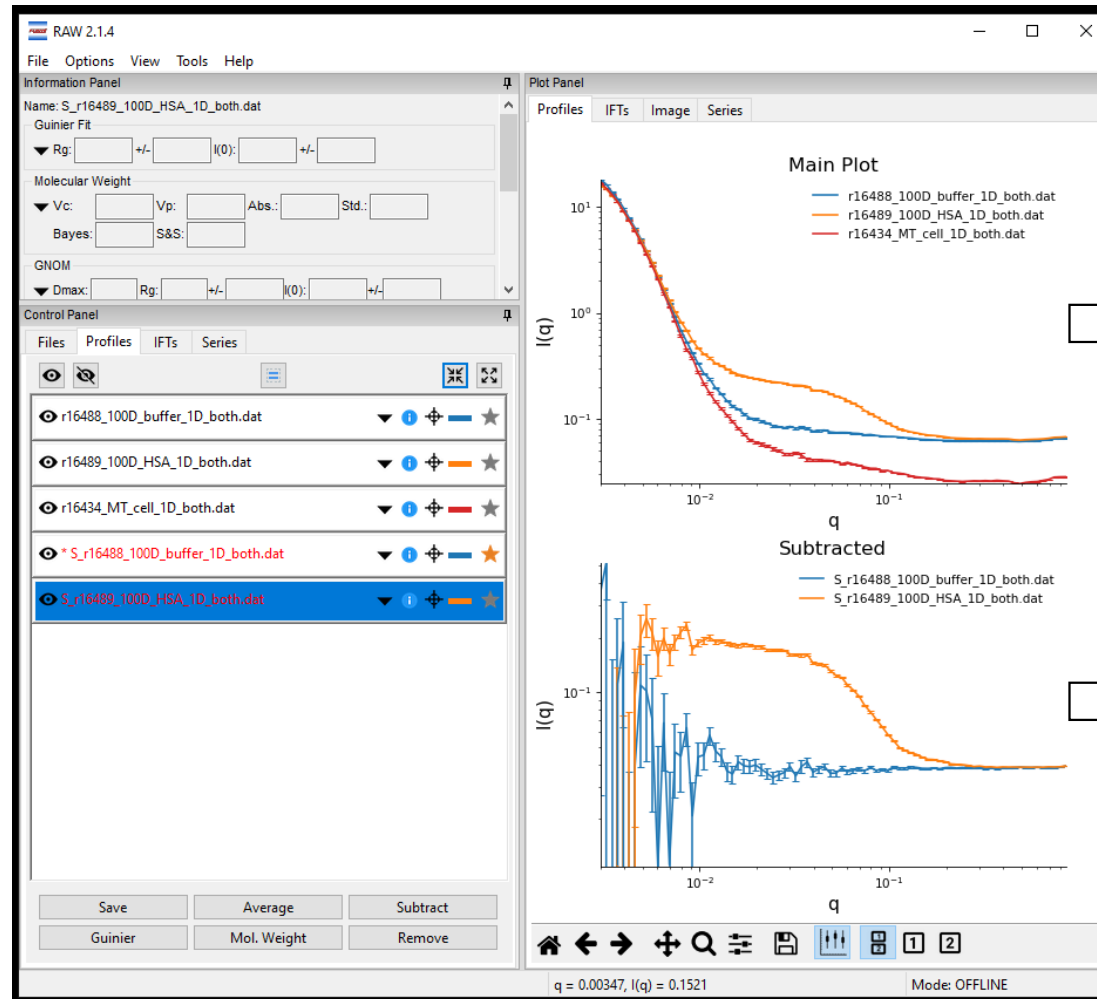
## Data manipulation

- Average
- Subtracted
- Merge
- Rebin

## Data collected

- Empty cell
- Buffer
- Sample

Empty cell and buffer are backgrounds



Before subtraction

After subtraction

# Background subtraction

## Second step - Buffer subtraction

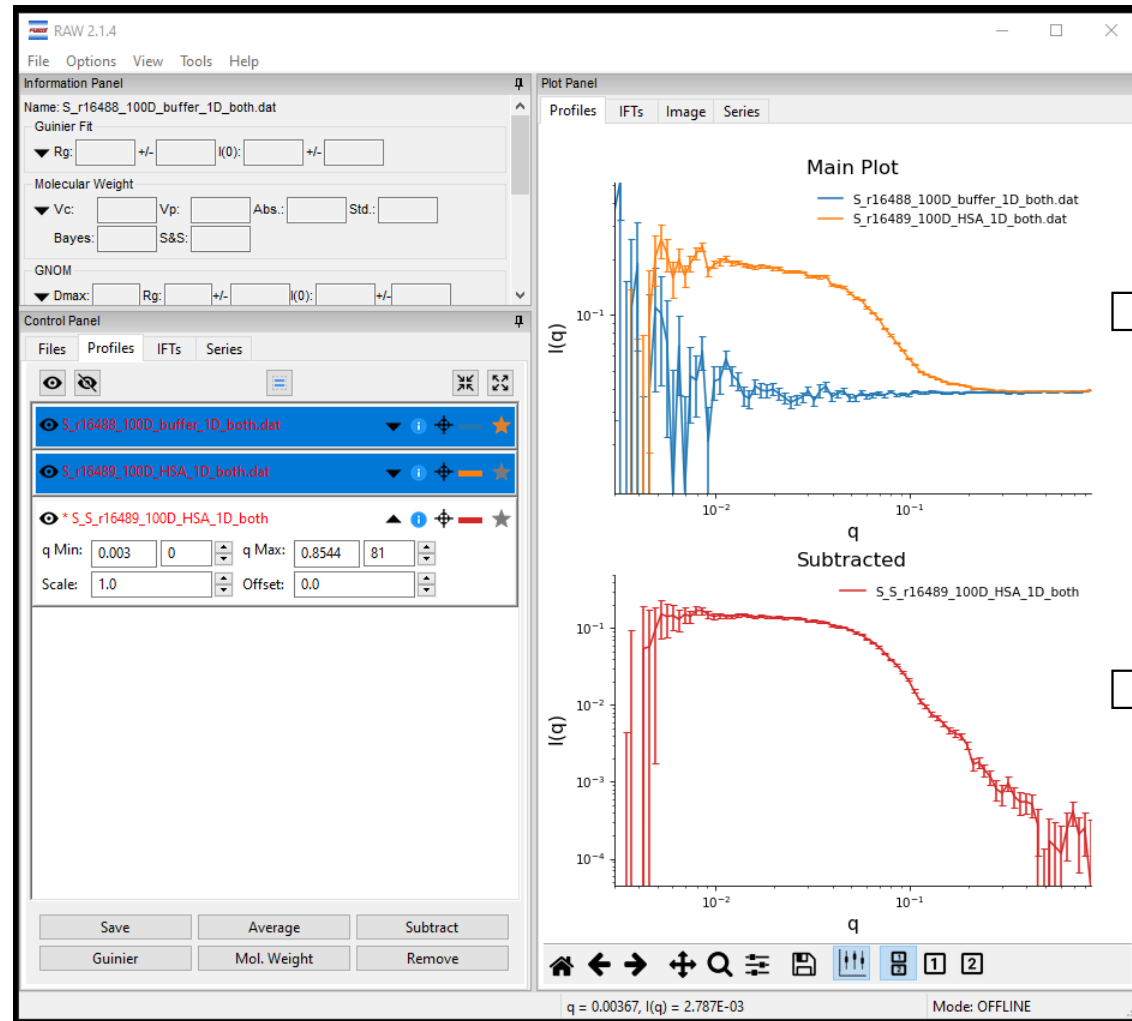
### Data manipulation

- Average
- Subtracted
- Merge
- Rebin

### Data collected

- Empty cell
- Buffer
- Sample

Empty cell and buffer are backgrounds



Before subtraction

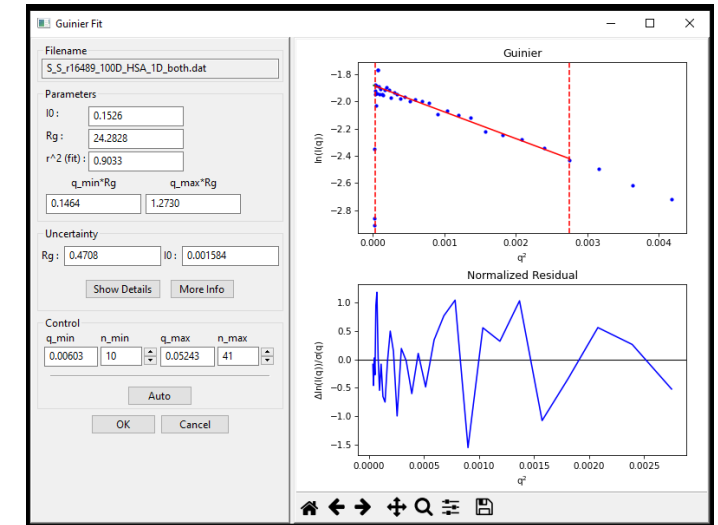
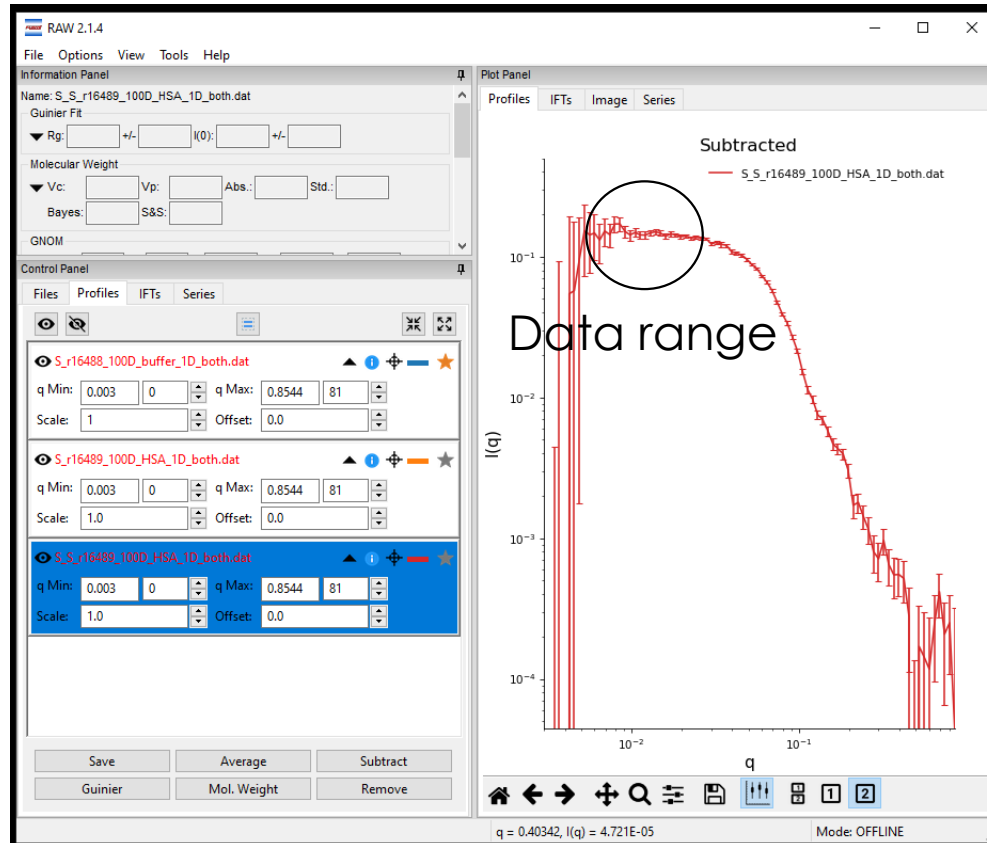
After subtraction

# Guinier Analysis

## Subtracted Scattering profile

## Guinier Analysis

- Subtract
- Average
- Weighted Average
- Other Operations >
- Remove
- Guinier fit
- Molecular weight
- IFT (BIFT)
- IFT (GNOM)
- Similarity Test
- Dimensionless Kratky Plot
- Other Analysis >
- Show image
- Show data
- Show header
- Show history
- Move to top plot
- Move to bottom plot
- Rename
- Save all analysis info
- Save item info
- Save selected file(s)
- Save report



## Guinier Analysis ( $R_g$ Guinier and $I_0$ Guinier)

$$I(q) \cong I(0) \exp\left(\frac{-R_g^2 q^2}{3}\right)$$

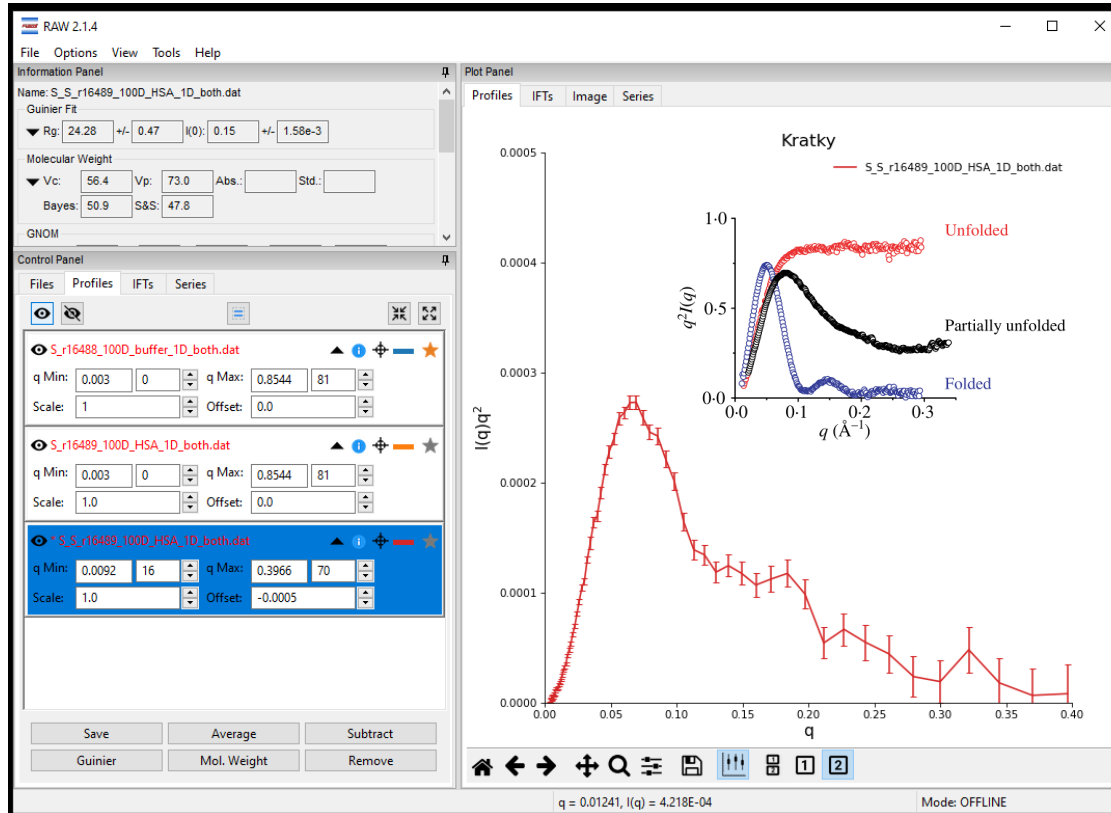
To get reliable Guinier plot /  $R_g$  analysis:

- $Q_{\min} R_g \leq 0.65$
- $Q_{\max} R_g \leq 1.3$  for globular
- $Q_{\max} R_g \leq 0.8$  for elongated
- $Q_{\max}/Q_{\min} > 2$

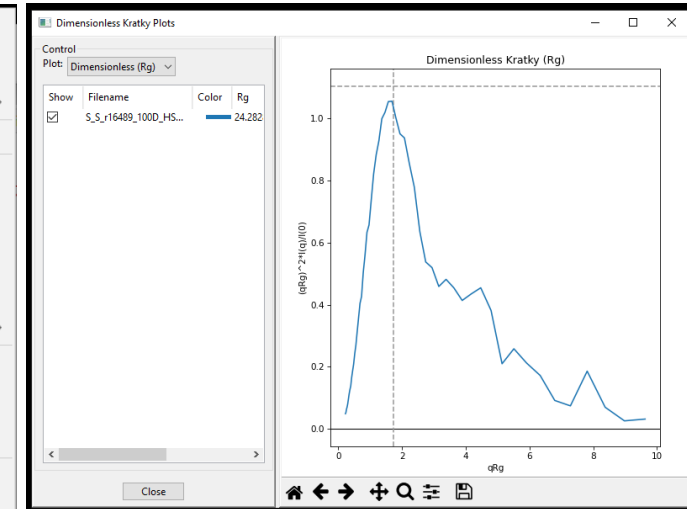
Guinier analysis allows model-free determination of the radius of gyration ( $R_g$  Guinier) and  $I_0$  Guinier (Forward scattering)

# Kratky plot analysis

## Kratky plot



## Dimensionless Kratky plot

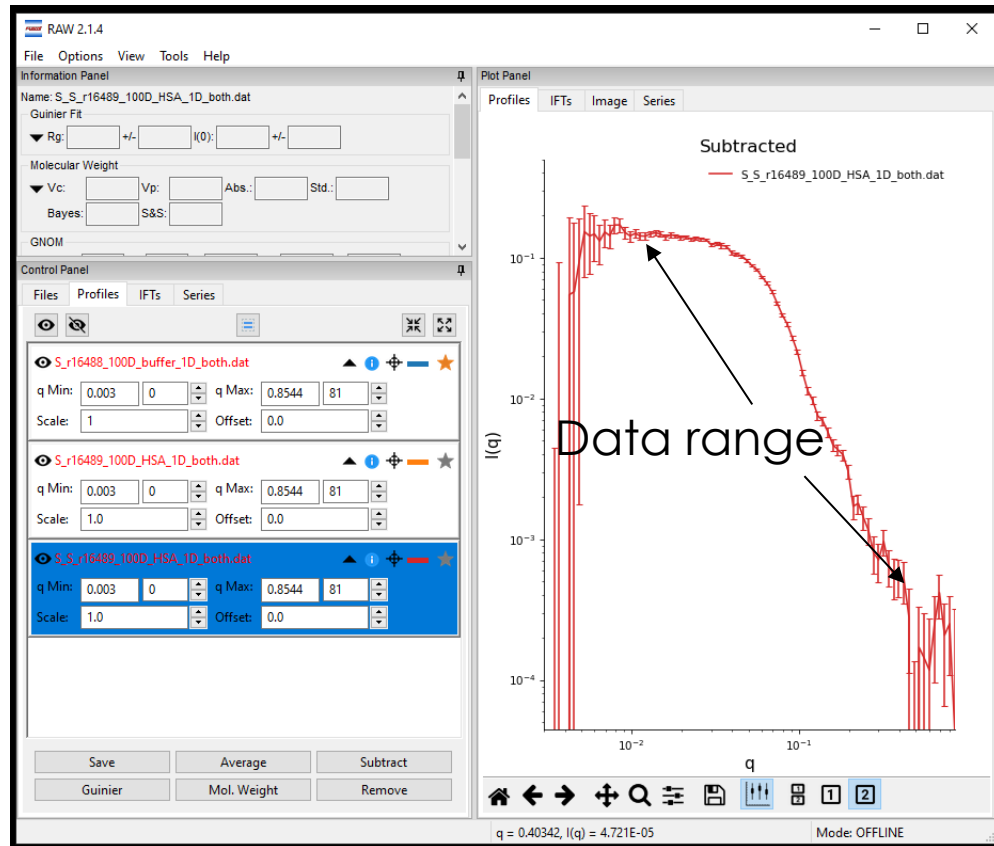


**Kratky Plot analysis:** A Kratky plot is a plot of  $I(Q) \times Q^2$  vs.  $Q$ . Kratky plot can qualitatively assess the flexibility and/or degree of unfolding in samples, as well as distinguish between extended to compact conformations.



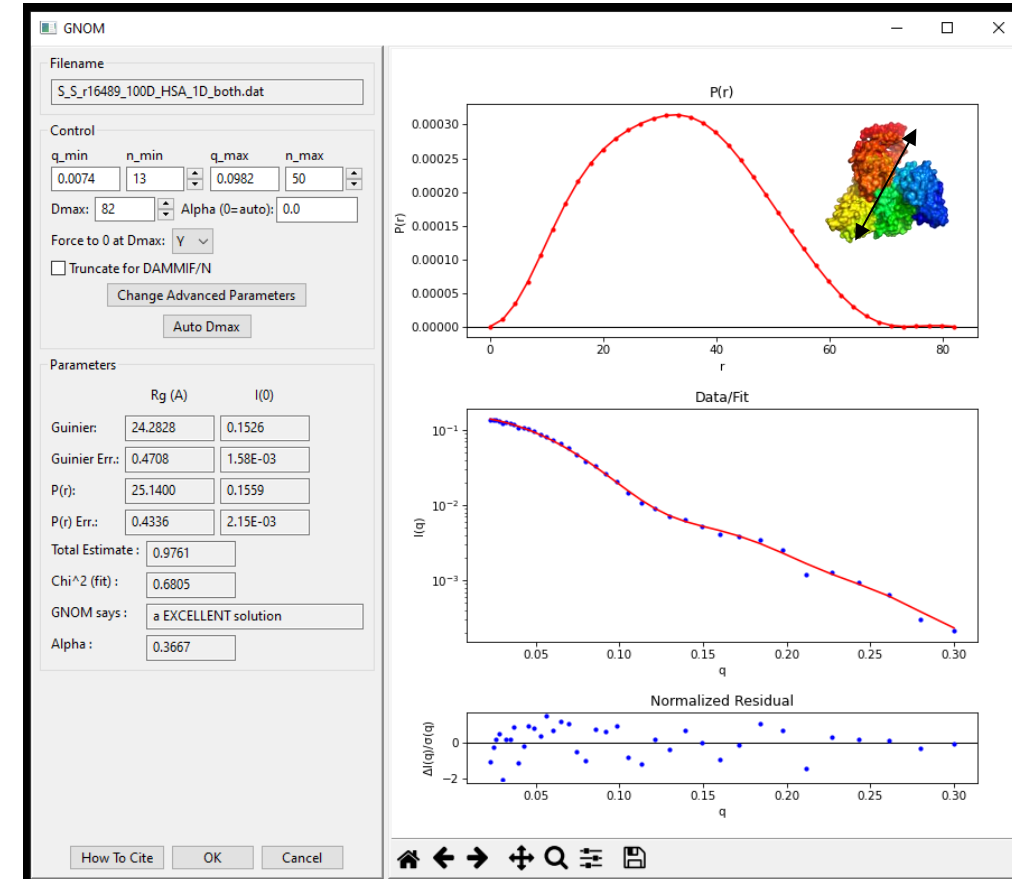
# Pair distance distribution

## Scattering profile



- Subtract
- Average
- Weighted Average
- Other Operations >
- Remove
- Guinier fit
- Molecular weight
- IFT (BIFT)
- IFT (GNOM)
- Similarity Test
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## P(r) profile

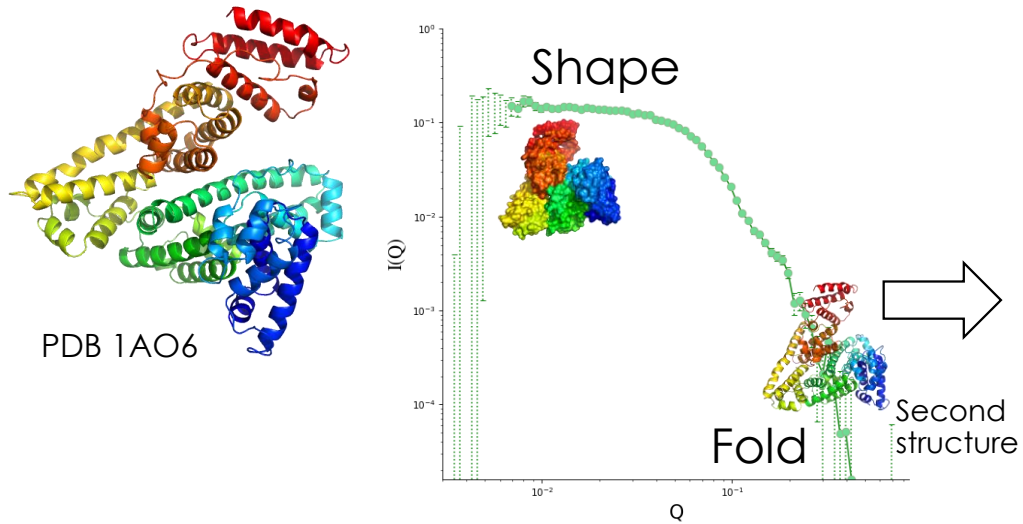


The Inverse Fourier Transform (IFT) transforms experimental (1D  $I(Q)$  vs.  $Q$ ) curves in reciprocal space to produce the pairwise distance distribution profile ( $P(r)$  vs.  $r$ ) of atoms in real space.

$P(r)$  analysis ( $D_{\max}$ ,  $R_g^{\text{GNOM}}$ , and  $I_0^{\text{GNOM}}$ )

# Comparison with PDB structures

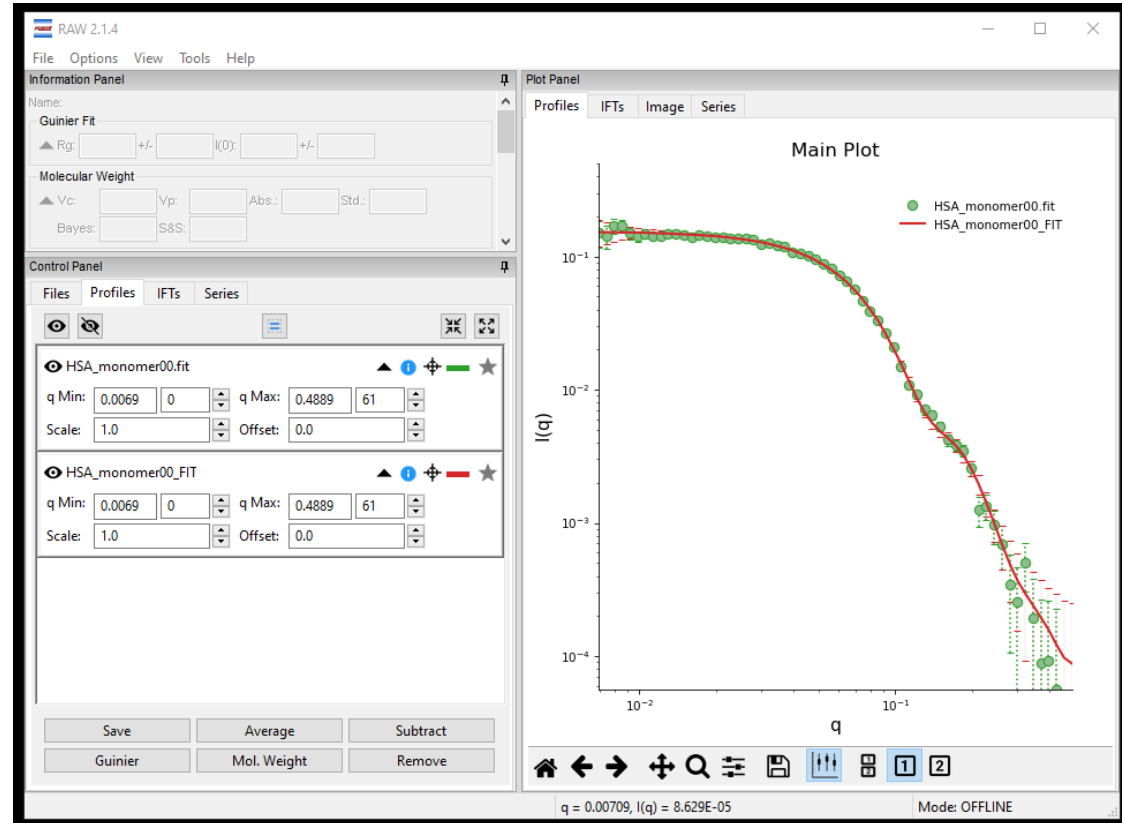
Crystal structure of Human Serum Albumin (HSA)



- Crystal structure
- Cryo-EM
- NMR
- AlphaFold
- Homology models

Created by CRYSON v. 27 on 07-Sep-2022 21:33:44

Data file name .....  
Model: HSA\_monome Rg: 25.16 Run: 00 Chi^2: 0.85



CRYSON is a program for evaluating the solution scattering from macromolecules with known atomic structure and fitting it to experimental scattering curves from Small-Angle Neutron Scattering (SANS).

# Molecular mass calculation

## Molecular mass calculation for monodispersed species

Standard equation from SAS

$$MW = \frac{I(0)N_A}{C_w(\Delta\rho v)^2}$$

$I(0)$  = Forward scattering

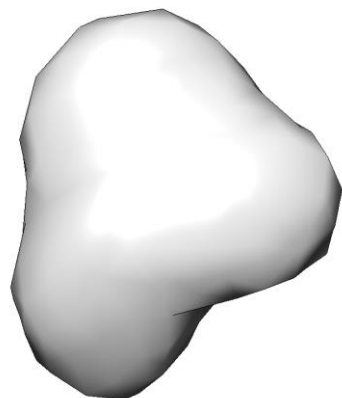
- Guinier Analysis
- Pair distance distribution

Intensity at Q=0	$I(0)$	0.16	$\text{cm}^{-1}$
Avogadro Number	$N_A$	6.023	$10^{23} \text{ mol}^{-1}$
Molar concentration	$C_w$	2.6	$10^{-3} \text{ g/cm}^3$
Scattering length Density			
Protein	rho_Prot	2.97	$\text{cm}^{-2}$
Solvent	rho_Solv	6.36	$\text{cm}^{-2}$
Difference	delta_rho	3.39	$\text{cm}^{-2}$
Partial specific volume	$v$	0.708	$\text{cm}^3/\text{g}$
Numerator	$I(0) N_A$	0.964	$\text{cm}^{-1} \text{ mol}^{-1}$
Denominator	$C_w (\text{delta\_rho } v)^2$	14.977	$\text{g}^{-1} \text{ cm}^{-1}$
Result	$M_w$	64.3	kDa

% D2O	Water	hProtein
0	-5.61E+09	1.94E+10
2	-4.21E+09	1.96E+10
4	-2.82E+09	1.99E+10
6	-1.43E+09	2.01E+10
8	-3.54E+07	2.03E+10
10	1.36E+09	2.05E+10
12	2.75E+09	2.07E+10
14	4.14E+09	2.09E+10
16	5.53E+09	2.11E+10
18	6.93E+09	2.13E+10
20	8.32E+09	2.15E+10
22	9.71E+09	2.17E+10
24	1.11E+10	2.19E+10
26	1.25E+10	2.21E+10
28	1.39E+10	2.23E+10
30	1.53E+10	2.25E+10
32	1.67E+10	2.27E+10
34	1.81E+10	2.29E+10
36	1.95E+10	2.31E+10
38	2.09E+10	2.33E+10
40	2.22E+10	2.35E+10
42	2.36E+10	2.37E+10
44	2.50E+10	2.40E+10
46	2.64E+10	2.42E+10
48	2.78E+10	2.44E+10
50	2.92E+10	2.46E+10
52	3.06E+10	2.48E+10
54	3.20E+10	2.50E+10
56	3.34E+10	2.52E+10
58	3.48E+10	2.54E+10
60	3.62E+10	2.56E+10
62	3.76E+10	2.58E+10
64	3.90E+10	2.60E+10
66	4.03E+10	2.62E+10
68	4.17E+10	2.64E+10
70	4.31E+10	2.66E+10
72	4.45E+10	2.68E+10
74	4.59E+10	2.70E+10
76	4.73E+10	2.72E+10
78	4.87E+10	2.74E+10
80	5.01E+10	2.76E+10
82	5.15E+10	2.79E+10
84	5.29E+10	2.81E+10
85	5.36E+10	2.82E+10
86	5.43E+10	2.83E+10
88	5.57E+10	2.85E+10
90	5.71E+10	2.87E+10
92	5.85E+10	2.89E+10
94	5.98E+10	2.91E+10
96	6.12E+10	2.93E+10
98	6.26E+10	2.95E+10
100	6.40E+10	2.97E+10

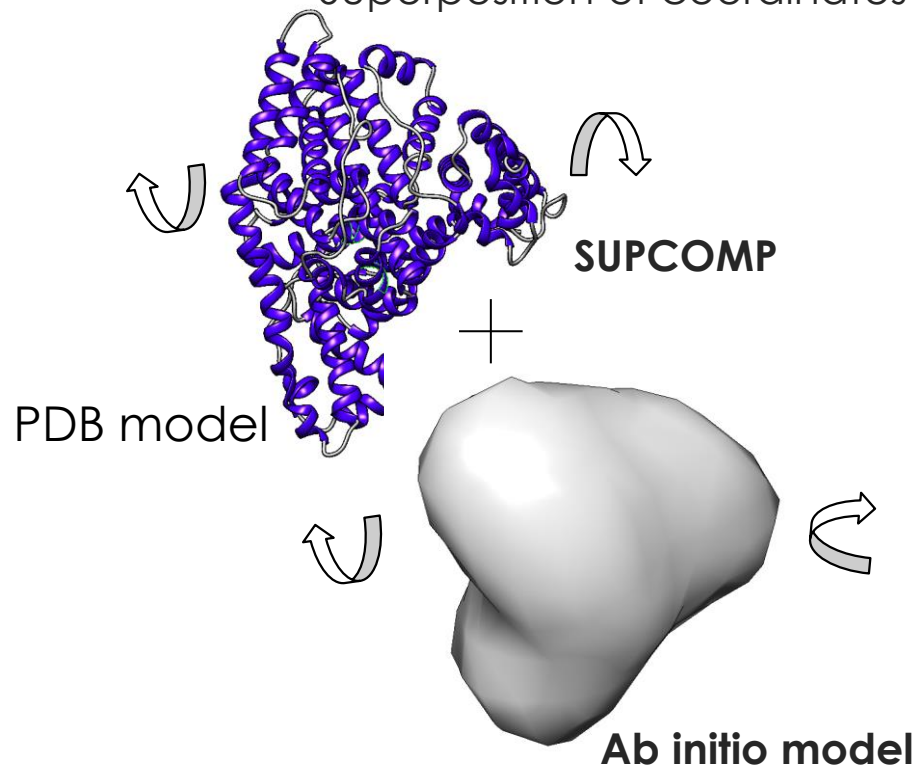
# Envelope of a macromolecule using Ab initio modeling

DAMMIN/DAMMIF

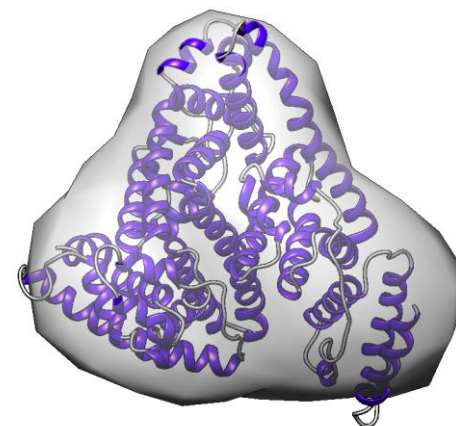


Ab initio model

Superposition of coordinates

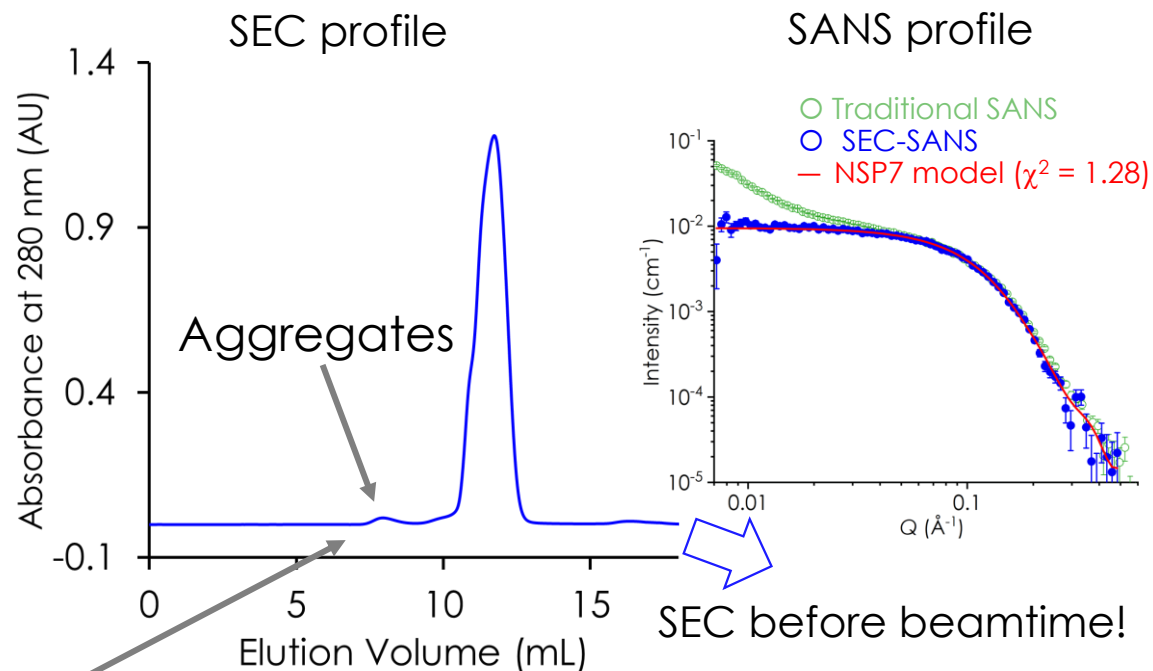


Overall shape or envelope



# Interparticle Interferences

## Attractive interaction between proteins in solution



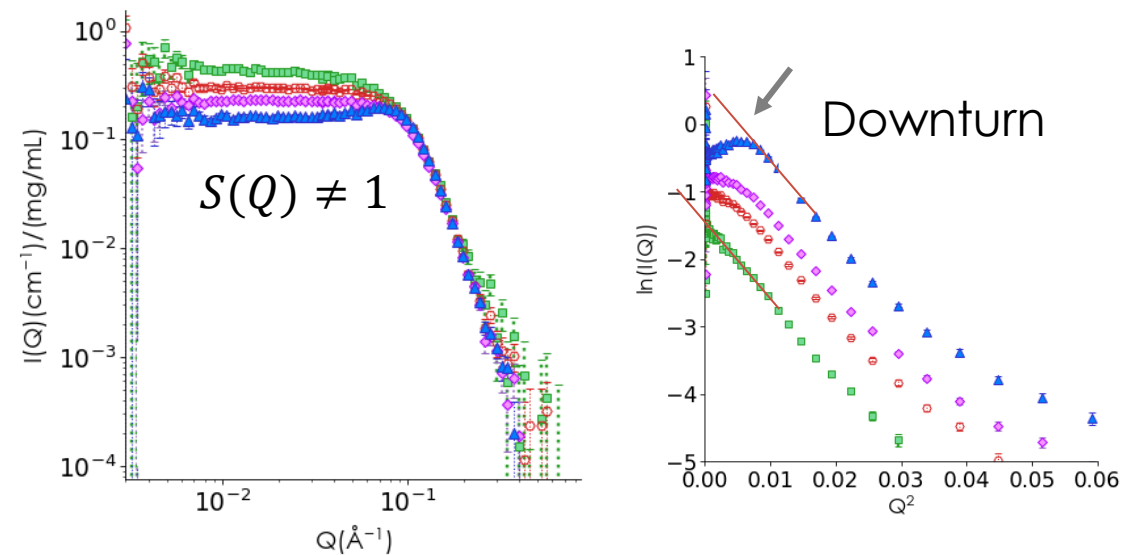
Note: Aggregates have low absorbance at 280 nm, but have high light scattering (MALS)

### Experiment details:

- Unlabeled Non-Structural Protein 7 (9.2 kDa)
- Injected ~500 μL at ~20 mg/mL
- 100% D<sub>2</sub>O buffer
- Superdex 75 10/300 GL (24 mL CV)
- Data collection triggered manually
- Higher order aggregates removed!
- Nsp7 exists as a dimer (18.4 kDa) in solution

## Repulsion interaction between proteins in solution

### SANS profile and Guinier Analysis



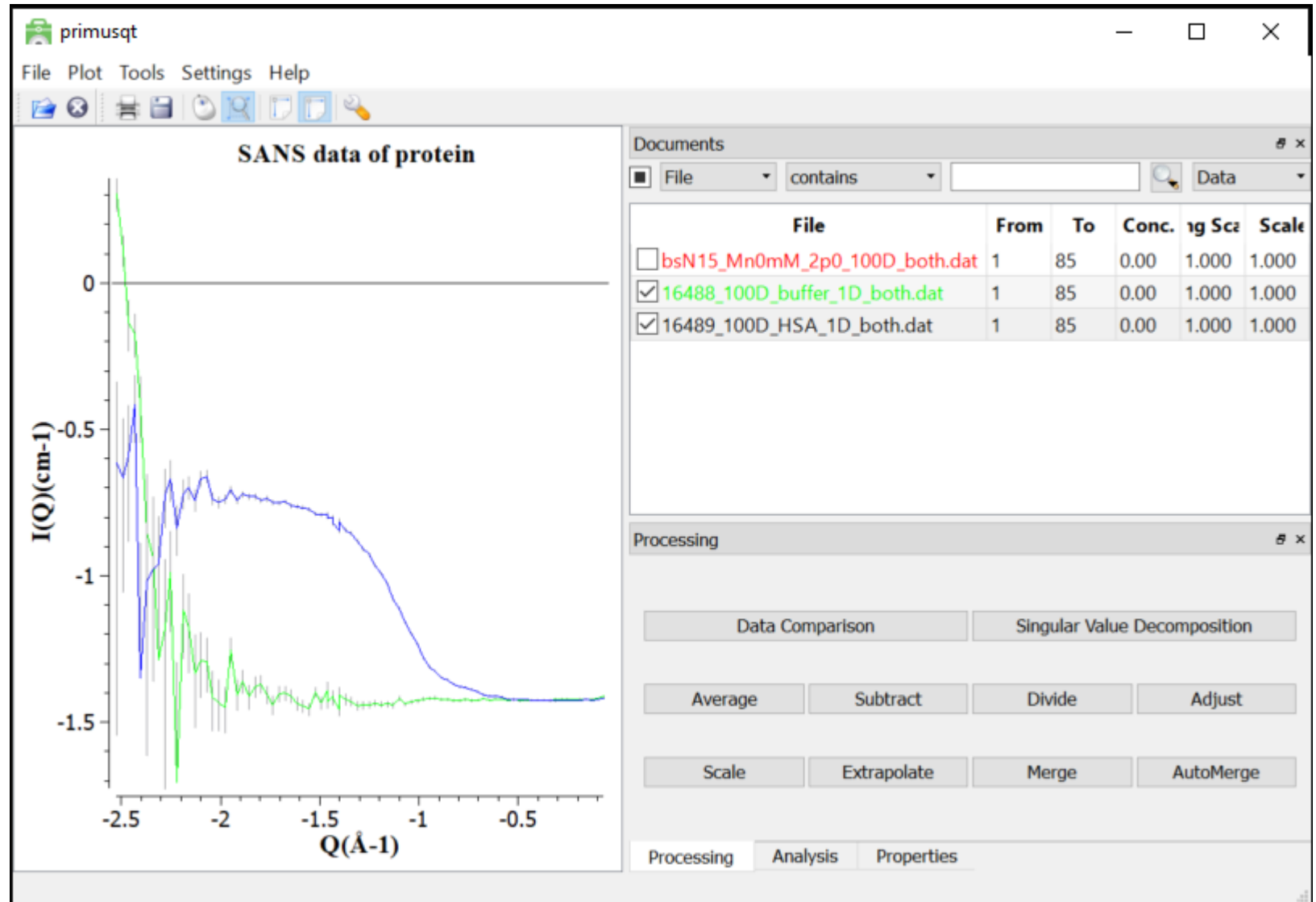
- Membrane protein in micelles
- Sample concentration was reduced 8-fold to reduce attractive interactions

# Resources

Data analysis software ATSAS  
3.1.1 - <https://www.embl-hamburg.de/biosaxs/software.html>

## Experimental data processing

- PRIMUS - GUI for manipulations and primary analysis of experimental 1D SAS data

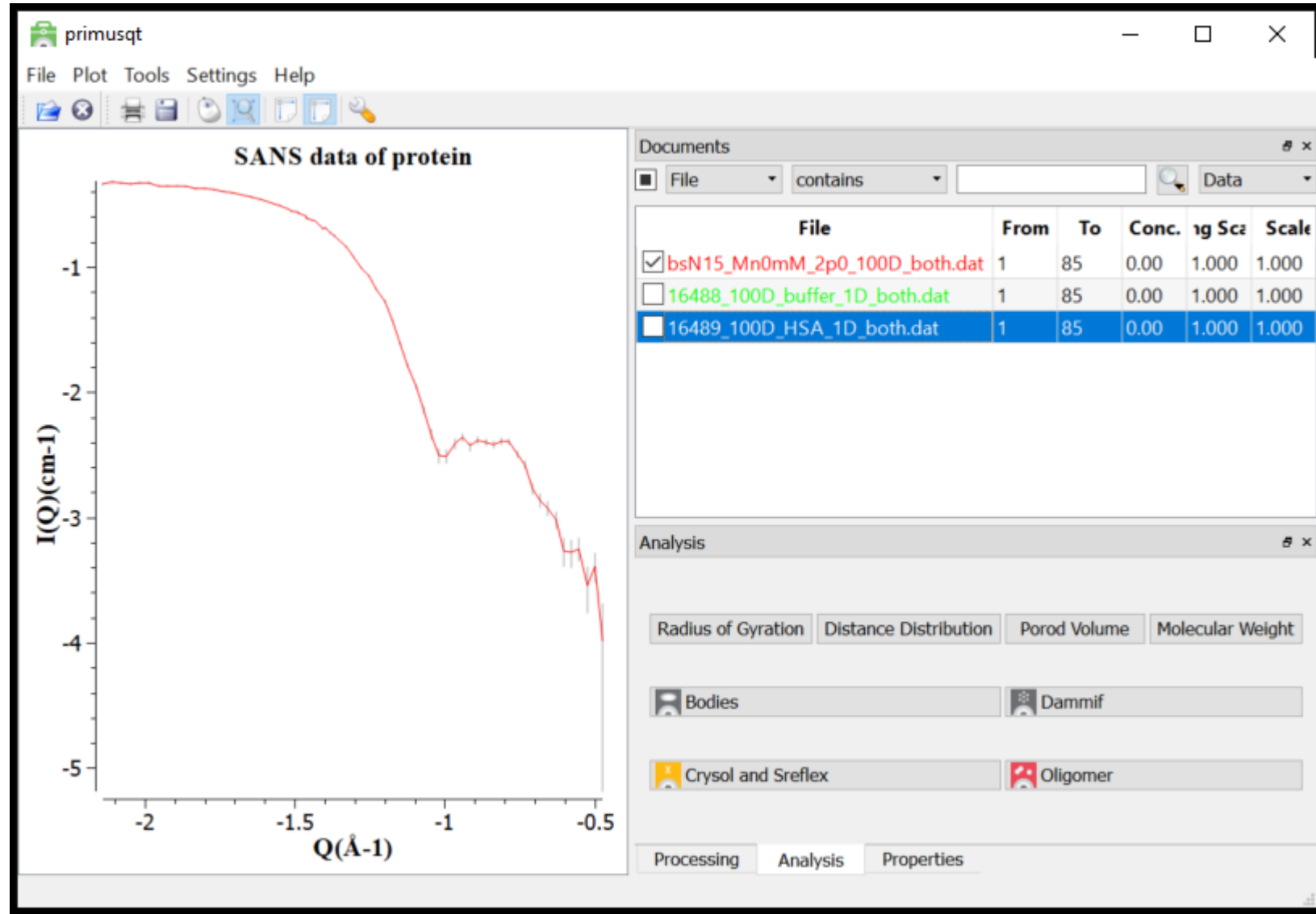


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# Facility Acknowledgment Statement

- A portion of neutron scattering research presented as examples in this introduction used resources at the High Flux Isotope Reactor or Spallation Neutron Source, DOE Office of Science User Facilities, operated by the Oak Ridge National Laboratory.
- The Bio-SANS of the Center for Structural Molecular Biology at the High Flux Isotope Reactor is supported by the Office of Biological and Environmental Research of the U.S. DOE.