

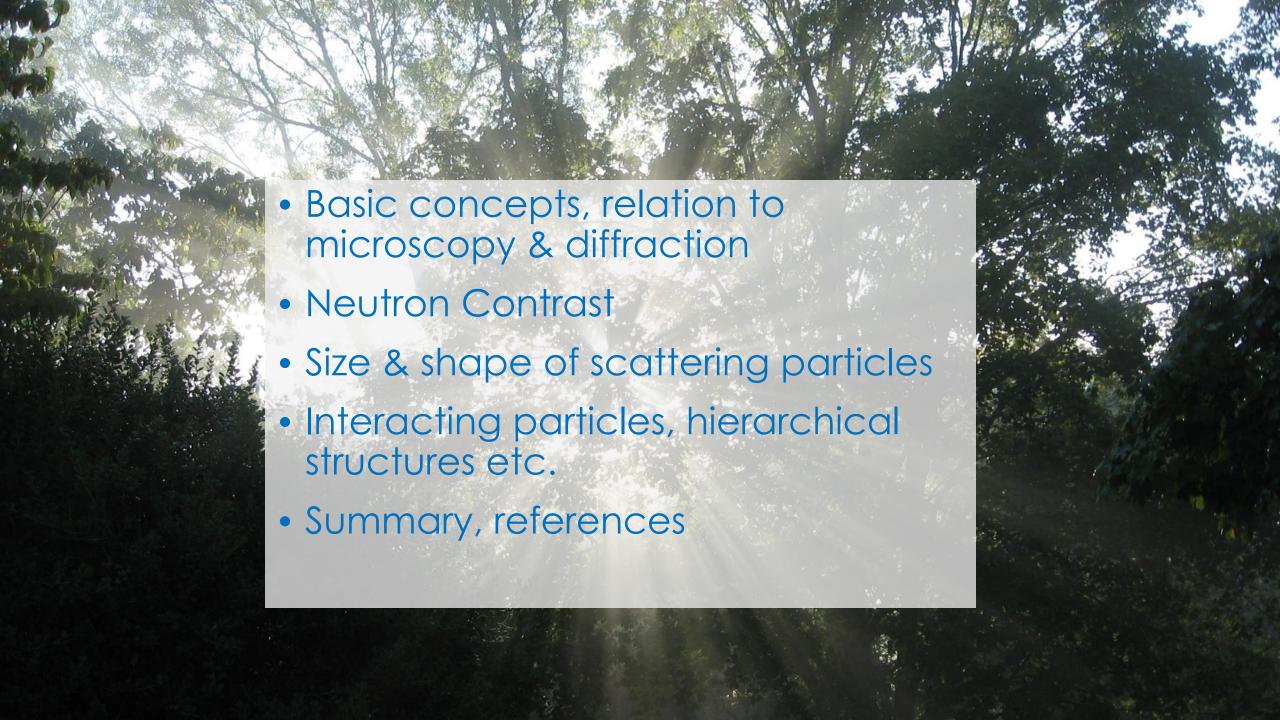
Introduction to Small-Angle Neutron Scattering

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Practical Guide for Biological Small-Angle Neutron Scattering Experiments and Data Analysis October 9-10, 2024

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Small Angle Scattering and Microscopy

Common features

- Size range 1nm-1μm
- Contrast labeling options (stains / isotope labels)

SANS practical aspects

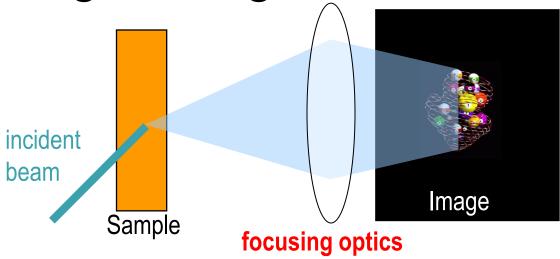
- No special sample preparation necessary (such as cryo freezing)
- Sample environments available for pressure, temperature, magnetic field etc.
- Non-invasive
- In-situ, time-resolved

Fundamental difference

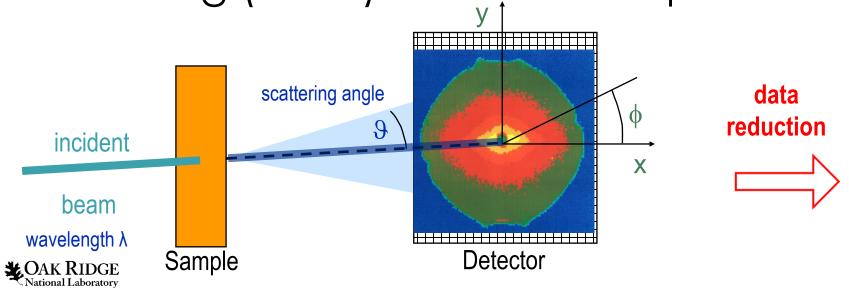
- "Real space" image with certain resolution
- Scattering pattern, averaged over volume
- Complementarity

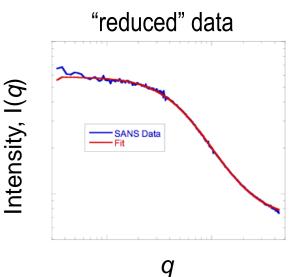


Microscopy: enlarged image



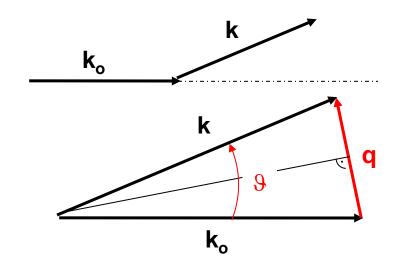
Scattering (SANS): interference pattern





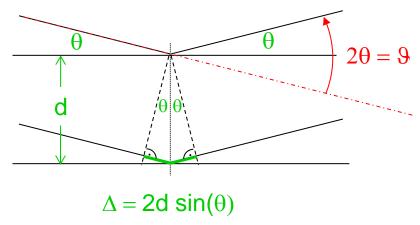
From scattering angle to scattering vector

Wave vector **k**: $|\mathbf{k}| = \mathbf{k} = 2\pi/\lambda$



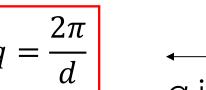
$$q = 2k \sin\left(\frac{\theta}{2}\right) = \frac{4\pi}{\lambda} \sin\left(\frac{\theta}{2}\right)$$

Bragg: waves with wavelength λ reflected by sets of lattice planes



if $\Delta = n \lambda$ then reflection, else extinction

$$\frac{1}{d} = \frac{2}{\lambda} \sin\left(\frac{\vartheta}{2}\right)$$



q in nm⁻¹ or Å⁻¹

scattering 'vector', a

also known as momentum transfer Q, q, h, k, s



Scattering and Diffraction (Crystallography)

- Diffraction from crystals, Scattering from anything else (less ordered)
- Same basic physics: interactions of radiation with matter
 - SAXS/WAXS, SAND/WAND
 - Instruments: resolution vs flux tradeoff (diffraction/scattering)
 - Diffraction needs crystal lattices, scattering does not.
 - Data analysis is very different in most cases.
- At small \mathbf{q} (small angles, large λ): observe nm-sized volume elements, "blobs" NOT atoms
 - Scattering length \rightarrow scattering length density **SLD**, symbol ρ
 - SA(N)S is sensitive to spatial non-uniformity of SLD:
 ΔSLD = Contrast → contrast variation!



Small Angle (Neutron) Scattering Intensity

• Interference of wavelets from distribution of nuclei (= structure) adds up to "net scattering" amplitude (Fourier transform of structure).

$$I(q) = \left| \int_{V} (\rho(\vec{r}) - \rho_s) e^{-i\vec{q} \cdot \vec{r}} d^3 r \right|^2$$

• Measured intensity I(q) is the magnitude square of amplitude.

$$I(q) = \underbrace{\frac{N}{V} (\Delta \rho)^2 V_p^2}_{\text{I(0)}} P(q) S(\vec{q}) \text{ where } P(q) = \left| F(q) \right|^2$$
structure form form factor factor amplitude

- I(q) also is the Fourier transform of the pair correlation function P(r).
- Incoming waves scatter off individual nuclei according to scattering length b (can be + or -).



Neutron Contrast – Atomic Scattering Lengths

Element	Neutrons (10 ⁻¹² cm)	X-rays (10 ⁻¹² cm)	Electr	ons
¹ H	-0.374	0.28	1	0
² H (D)	0.667	0.28	1	0
С	0.665	1.67	6	
N	0.940	1.97	7	
0	0.580	2.25	8	
Р	0.520	4.23	15	

For Small Angle Scattering

SANS (concept applies also to X-rays)

 $SL \rightarrow SLD \rightarrow \Delta SLD$

$$\overline{\rho} = \frac{1}{V} \sum_{i=1}^{N} b_i$$



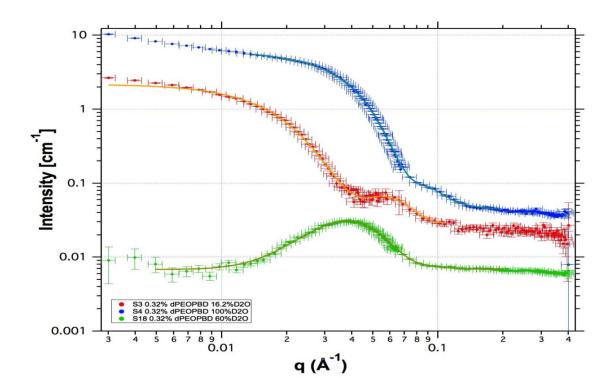


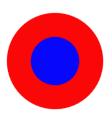


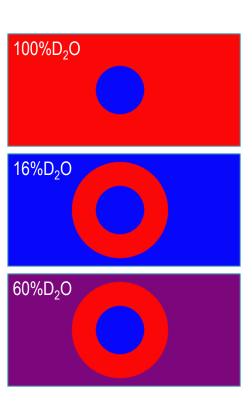
MOV

Increasing SAS information content by neutron contrast variation

- Block copolymer micelle with deuterated PEO block
- In solution neutrons probe:
 - Core only in 100% D₂O
 - Shell only in 16% D₂O / 84% H2O
 - Core-shell-interference term only in 60% D₂O / 40%H₂O



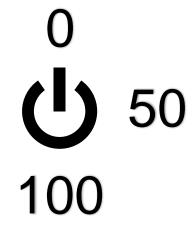




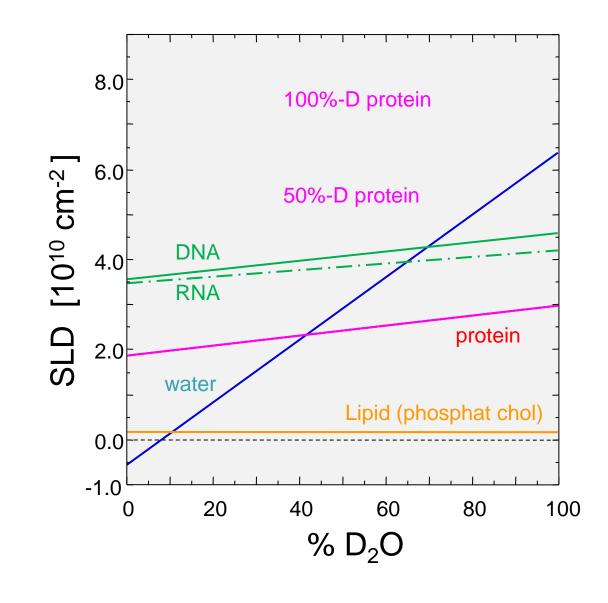


We can tune contrast by specific deuteration

D-labeling tuning knob



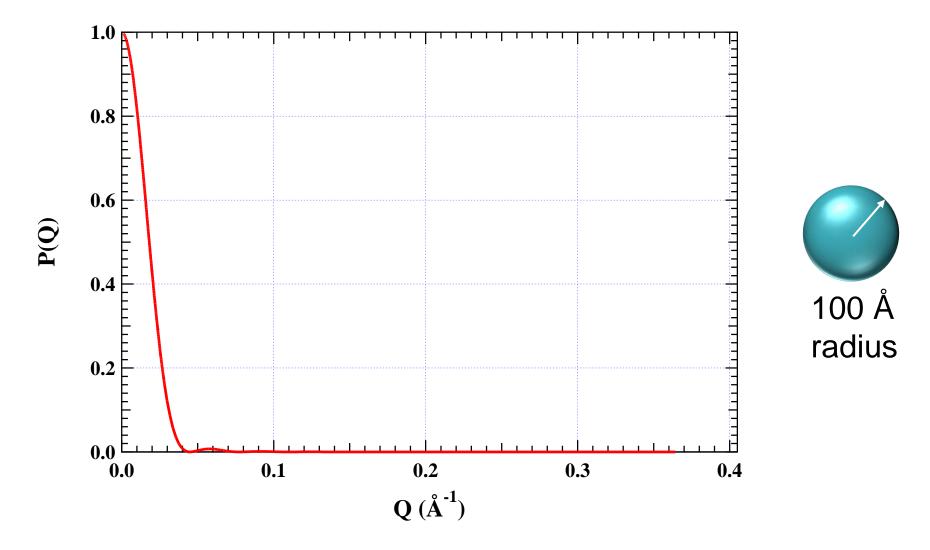
- 100% deuteration is not always what you want
- Proteins, polymers, organic molecules



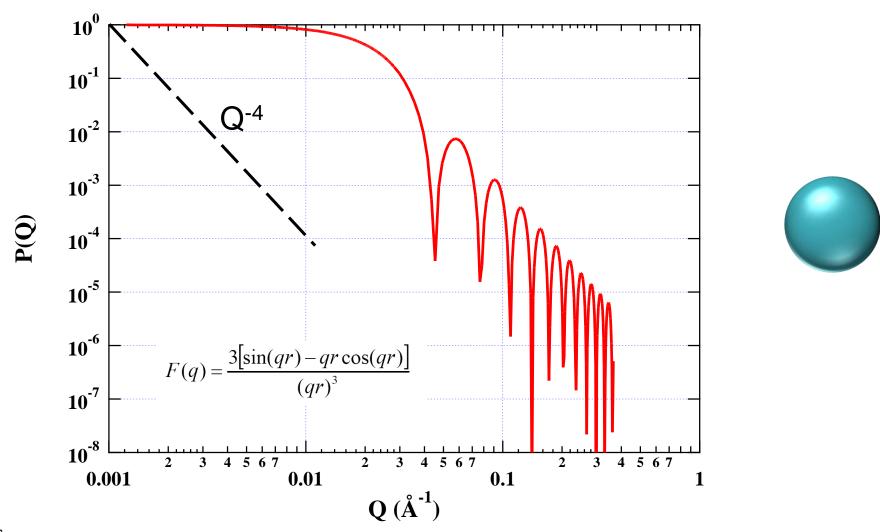


Sphere

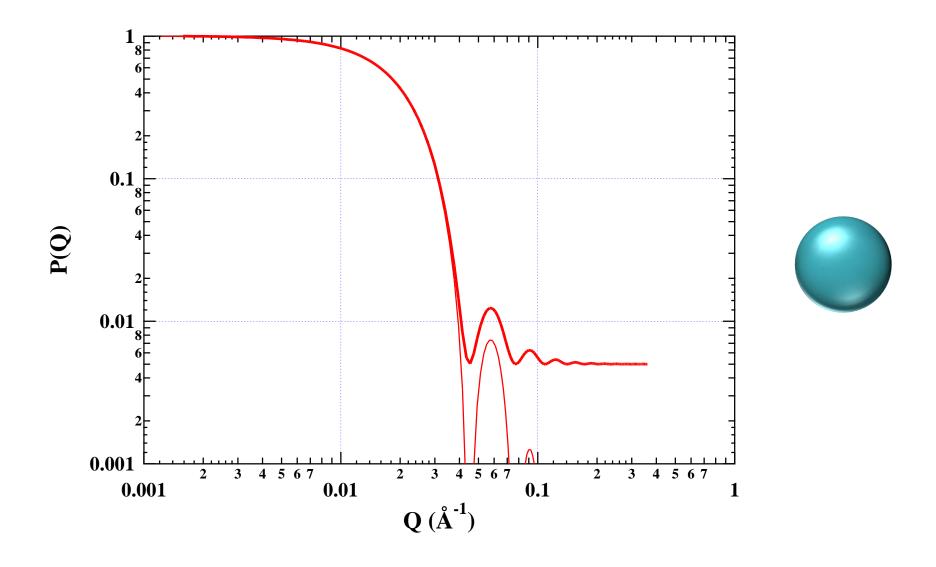
precisely: monodisperse sphere of uniform density with sharp and smooth surface



Sphere

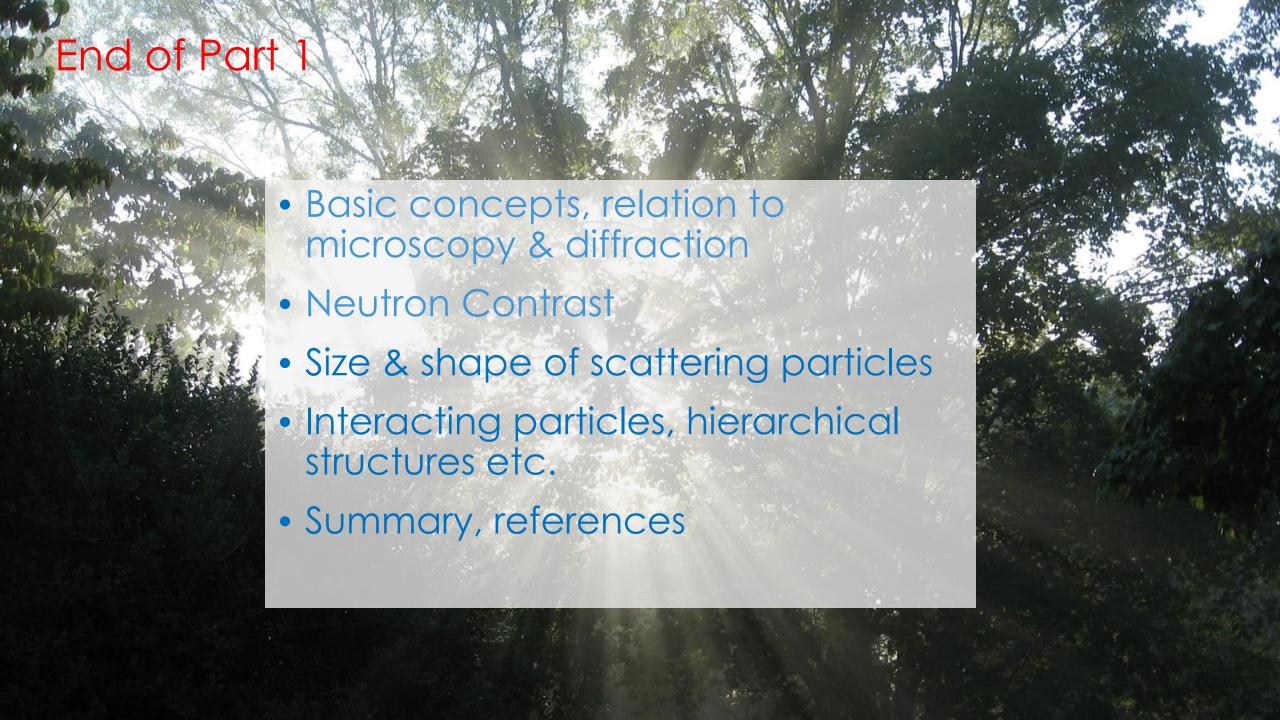


In practice: sphere + constant background

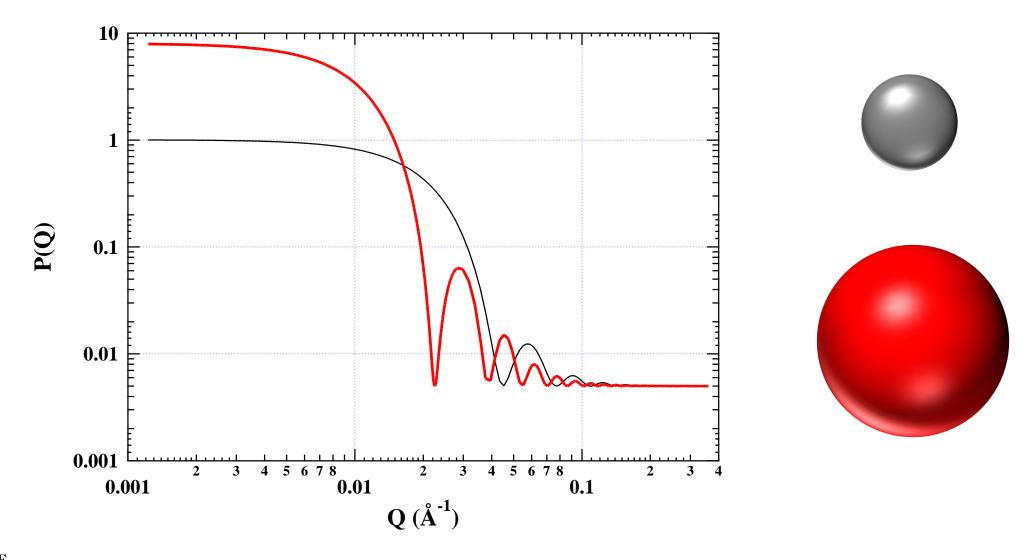


Some Notes on Background Subtraction

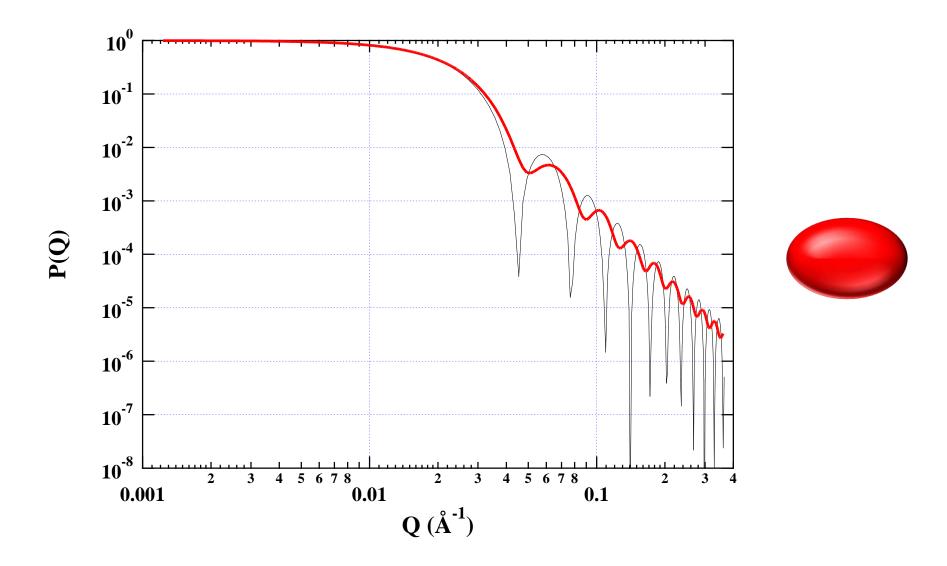
- A crucial step in data reduction/analysis
- Different sources of background (ambient, cell, intrinsic to sample) often require different treatments.
- Coherent scattering intensity drops over orders of magnitude with q (log scale), incoherent background is constant.
- More hydrogen means more incoherent background, this can influence the choice of contrast. (Less background is better.)



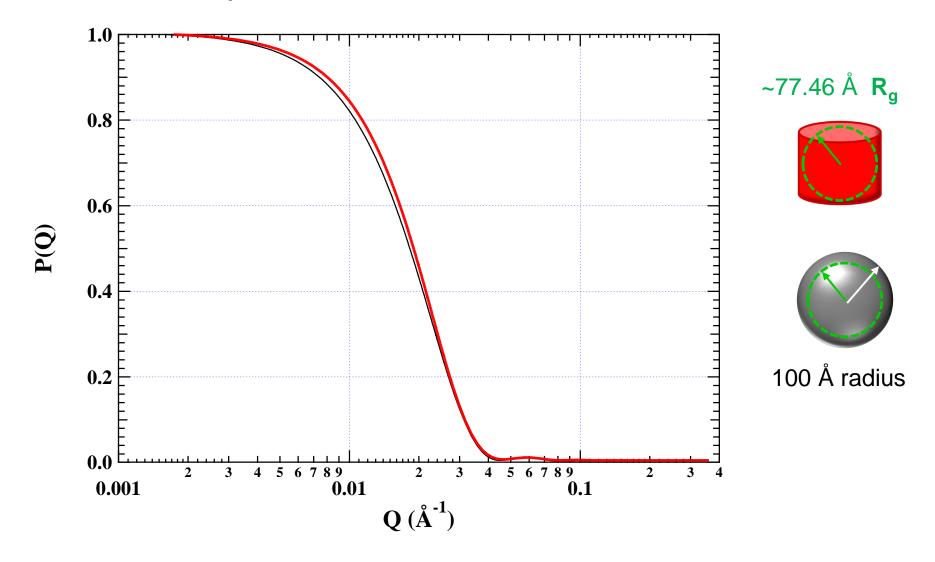
Spheres of different sizes



Ellipsoid aspect ratio 1.5



Circular Cylinder -with same Rg as the sphere





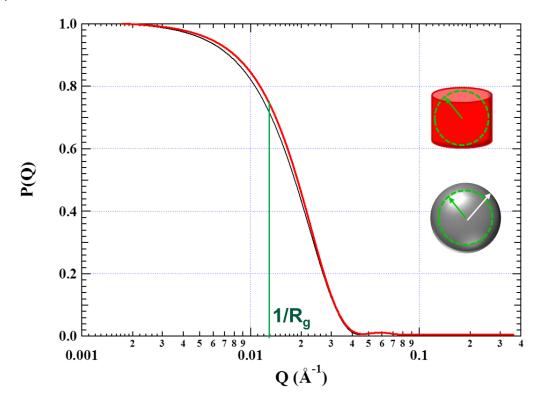


Guinier Analysis size of any kind of object

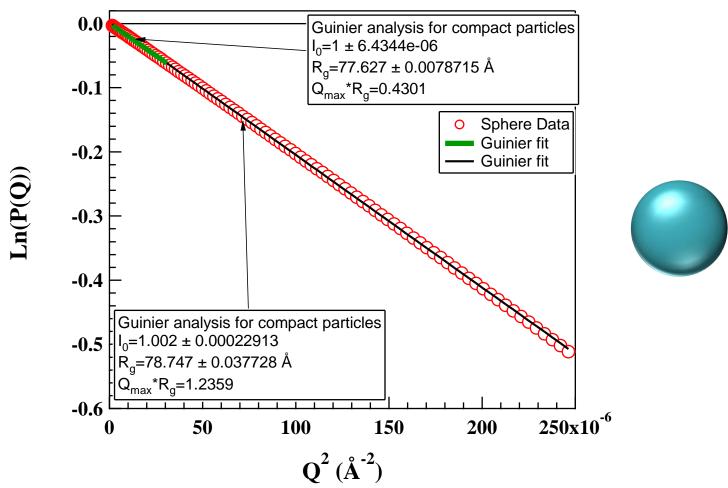
 At small enough Q anything that could reasonably be considered a discrete object follows Guinier's approximation.

$$\ln[I(q)] \propto q^2 R_g^2 / 3$$
 $qR_g < 1$; sphere : $R = \sqrt{\frac{5}{3}} R_g$

 Modified Guinier approximations exist to determine cross sectional radius of rods or thickness of sheets



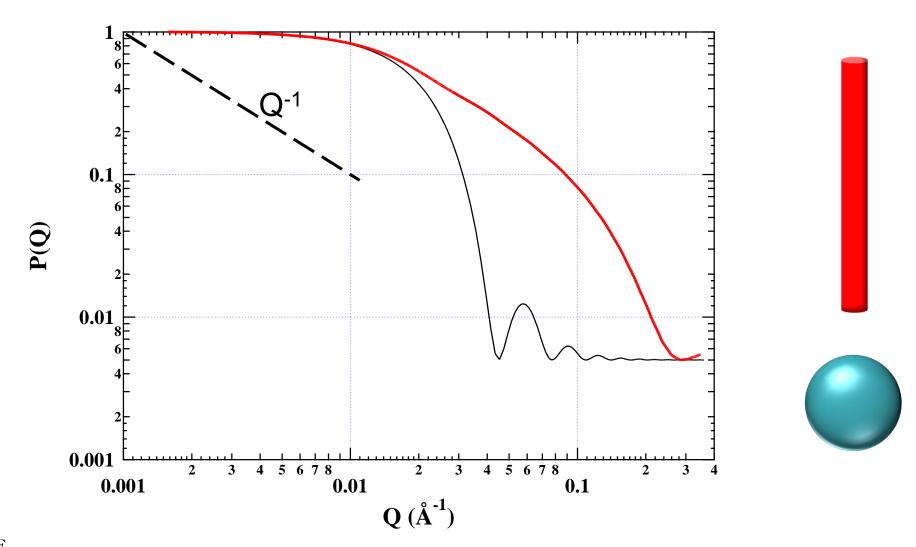
Guinier Analysis size of any kind of object



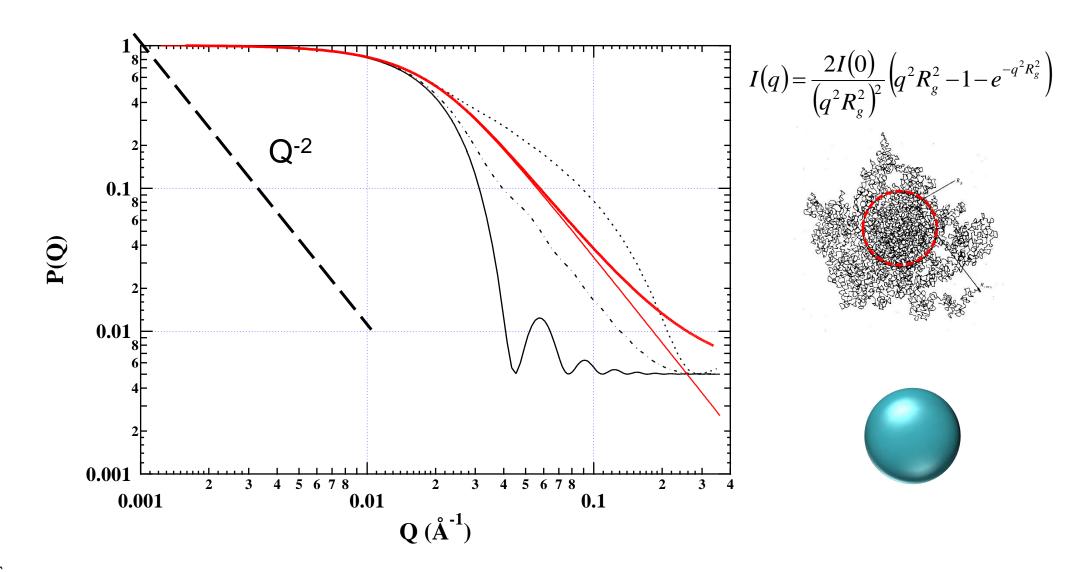
Precise R_g is 77.46 Å



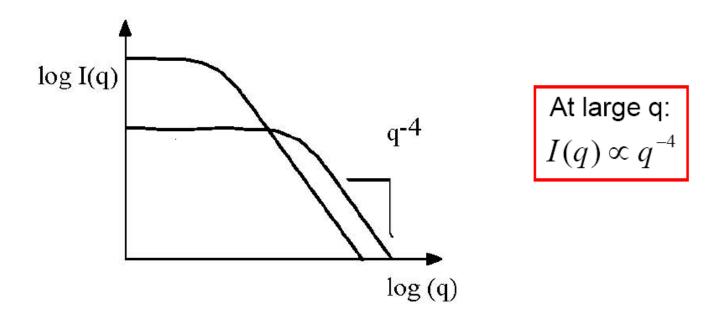
"Long & thin" cylinder



Polymer coil



Surface Scattering - Porod

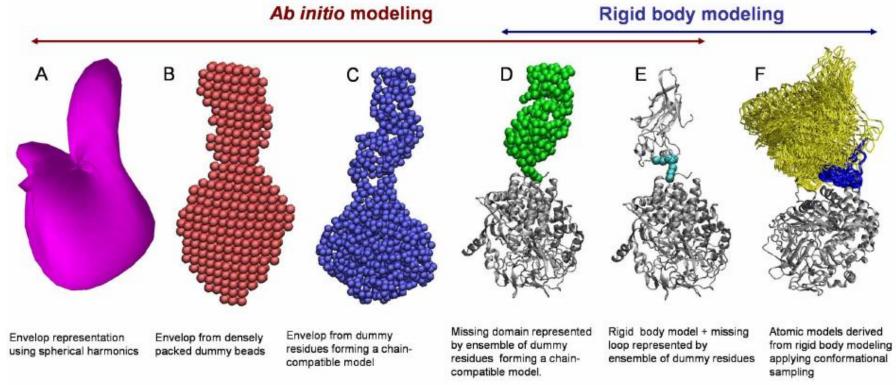


Specific Surface Area, S_V

$$\lim_{q \to \infty} I(q) = 2\pi S_V |\Delta \rho|^2 q^{-4}$$

But, fractal rough interfaces: Q^{-x} , 3 < x < 4Diffuse interfaces: x > 4

SAS Form Factor Modeling used in structural biology

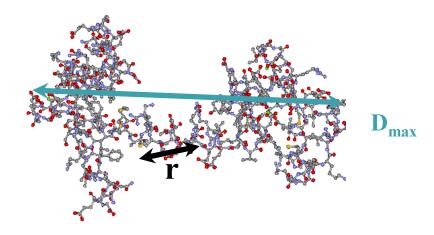


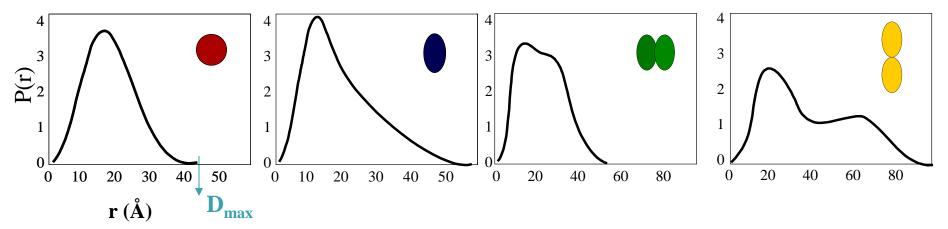
- Spherical Harmonics (Svergun, Stuhrmann, Grossman ...)
- Aggregates of Spheres (Svergun, Doniach, Chacón, Heller ...)
- Sets of High-resolution Structures (Svergun, Heller, Grishaev, Gabel ...)
- Simple Shapes and Custom Approaches (Henderson, Zhao, Gregurick, Heller ...)



Pair correlation function and shape

P(r): inverse Fourier transform of scattering function: Probability of finding a vector of length r between scattering centers within the scattering particle.





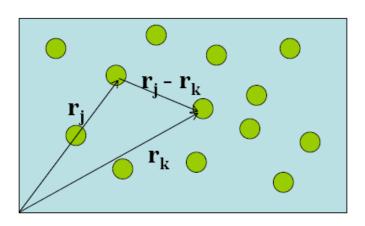
Shape: Modeled as a uniform density distribution that best fits the scattering data.



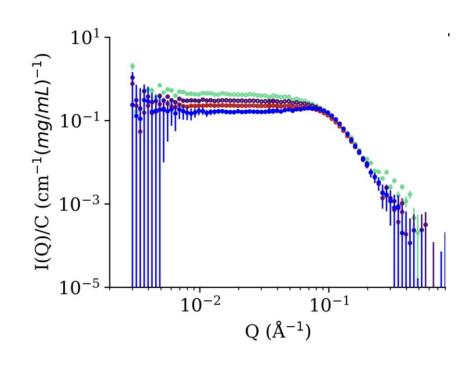
Interparticle Structure Factor S(Q)

$$I(q) = \frac{N}{V} (\Delta \rho)^2 V_p^2 P(q) S(\vec{q})$$

$$S(\vec{q}) = 1 + \left\langle \sum_{k=1}^{N} \sum_{\substack{j=1\\j \neq k}}^{N} e^{i\vec{q} \cdot (\vec{r}_k - \vec{r}_j)} \right\rangle$$



I(q) is modulated by interference effects between radiation scattered by different scattering bodies.

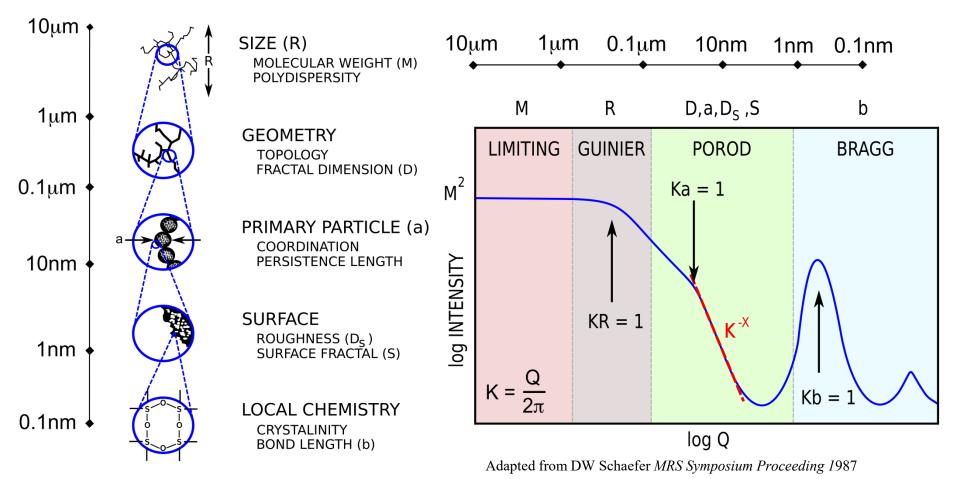


S(q) examples: hard sphere potential, sticky sphere, screened coulomb etc.

S(q)-P(q) is not always valid and useful!



Structural Hierarchy (particulate)

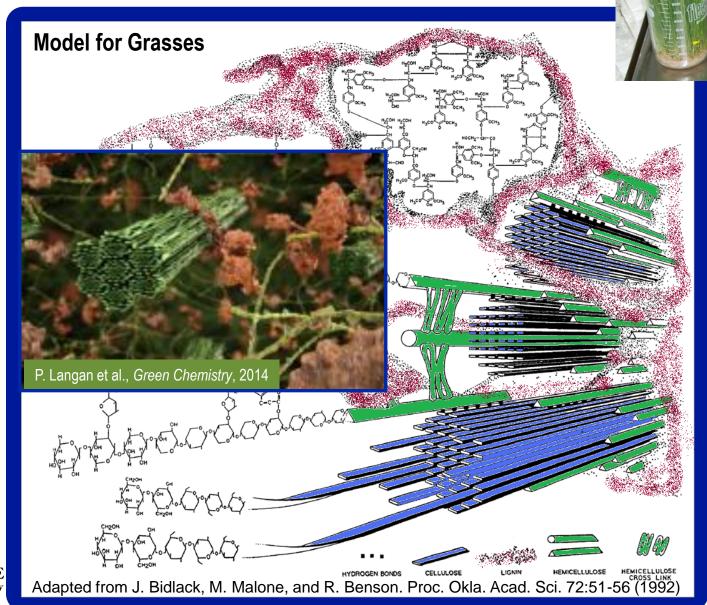


Structural information viewed on five length scales. Structural features at larger length scales are observed at smaller Q.

Scattering analysis that describes hierarchical structures: Mass Fractal (Teixeira), Unified Fit (Beaucage) combine power law scattering ranges with R_a transitions



"BIOMESS"



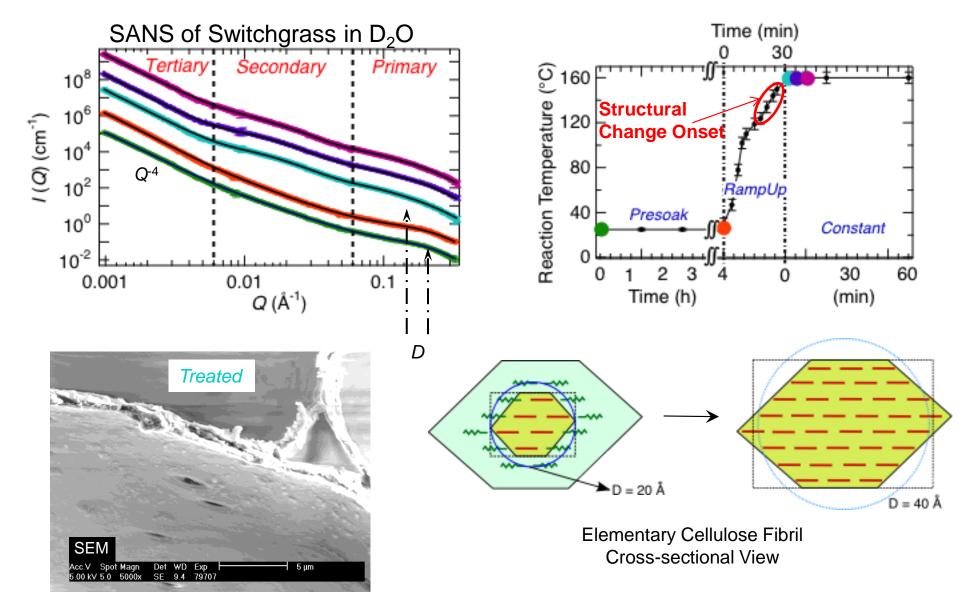


- Use D₂O for contrast
- Observe changes over time in pressure reaction cell with SANS

Outcome

New understanding of what processes actually happen during industrial pretreatment.

Dilute Acid Pretreatment of Switchgrass





SANS Summary

- Applications are in the nm to µm range and otherwise only limited by imagination.
- SANS does not see atoms, but larger, interesting features over many length scales.
- Precision of structural parameters such as $R_{\rm g}$ can be 1Å or better.
- SANS is used alone, but often complementary to other methods, such as microscopy, NMR.
- Scattering is similar to diffraction but does not require crystals.
- Data analysis is application dependent, using a diverse set of approximations, models and software.



SA(N)S Reference Suggestions

- Guinier, A., Fournet, G. 1955. Small-Angle Scattering of X-rays. John Wiley & Sons, New York.
 The classical work on small-angle scattering. Even though focused on x-rays, much of the theory and data interpretation apply just as well to neutrons.
- Pedersen, J. S., 1997. Analysis of small-angle scattering data from colloids and polymer solutions: modeling and least-squares fitting. Adv. Colloid Interface Sci. 70:171-210.
 Contains a comprehensive list of form factors and structure factors that are used for interpreting small-angle scattering data.
- Urban, V. S., 2012. Small-Angle Neutron Scattering. In: Characterization of Materials, edited by Elton N. Kaufmann. Copyright 2012 John Wiley & Sons, Inc.
 - A concise introduction to theory and practical considerations of Small-Angle Neutron Scattering.
- Chaudhuri, Muñoz, Qian, Urban (Editors), 2017. Biological Small Angle Scattering: Techniques, Strategies and Tips. Springer.

SA(N)S References continued

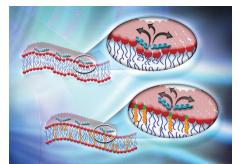
- Roe, R. J. 2000. Methods of X-Ray and Neutron Scattering in Polymer Science. Oxford University Press, New York and Oxford.
 - Even though focused on polymers, this book gives a very thorough account on the basic scientific principles of small-angle scattering in a fashion that is accessible to non-expert scatterers.
- Higgins, J. S., and Benoît, H. C. 1994. Neutron Scattering from Polymers. Clarendon Press, Oxford.
 A comprehensive description on neutron scattering and in particular small angle neutron scattering. Even though focused on polymers, the book is very useful for anyone interested in small angle neutron scattering.
- Lectures from the National School on Neutron and X-Ray Scattering: http://neutrons.ornl.gov/nxs
- PROBING NANOSCALE STRUCTURES THE SANS TOOLBOX, by Boualem Hammouda (NIST): https://www.ncnr.nist.gov/staff/hammouda/the_SANS_toolbox.pdf
- DOE BER Structural Biology Portal https://berstructuralbioportal.org/

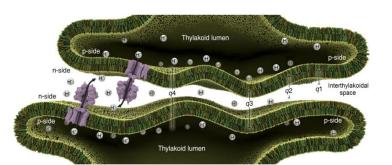


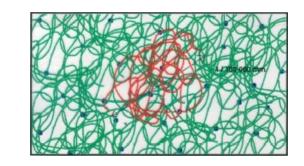


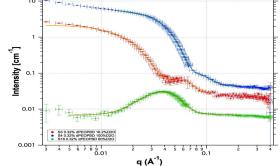
Cases for using neutron contrast

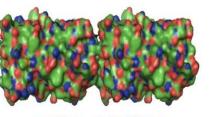
- 1. Zero (0) natural electron density contrast
- 2. Reducing degeneracy / increasing information content of SAS data
- 3. Overcoming extremely low signal to noise "needles in haystacks"
- 4. Zooming in on specific features in very complex systems biomass, live cells
- 5. Combination with other valuable neutron traits: fine energy resolution, no radiation damage, high penetration/in situ













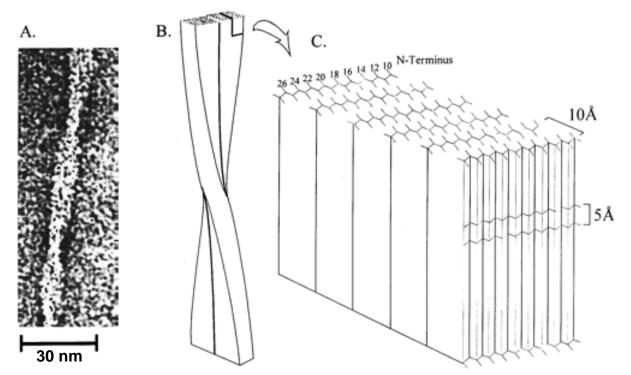






Complementary methods example: β-Amyloid

- Alzheimer's DiseaseAmong leading causes of death
- Miss-folded peptides form hierarchical ordered fibril structures & plaques
- Structure established using synthetic model peptides and complementary methods NMR, SANS, EM



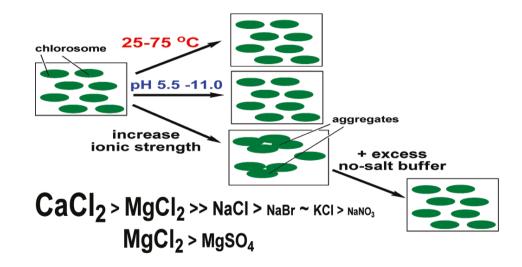
T.S. Burkoth et al. J. Am. Chem. Soc. 2000, 122, 7883-7889

- NMR
 - $-\beta$ -fold
- SANS
 - Fiber shape
 - Diameter
 - 6 sheet stack
- EM
 - Overall morphology
 - Twist



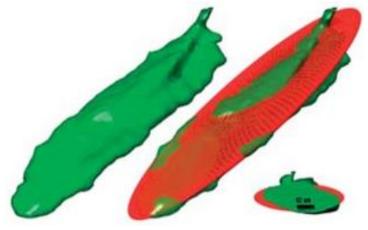
Ellipsoid example: Chlorosomes stable under harsh conditions including entrapment in silica sol-gel

Neutron Scattering probes chlorosome under range of temperature, pH and salt conditions



Remarkable stability of the chlorosome, reversible association in high ionic strength

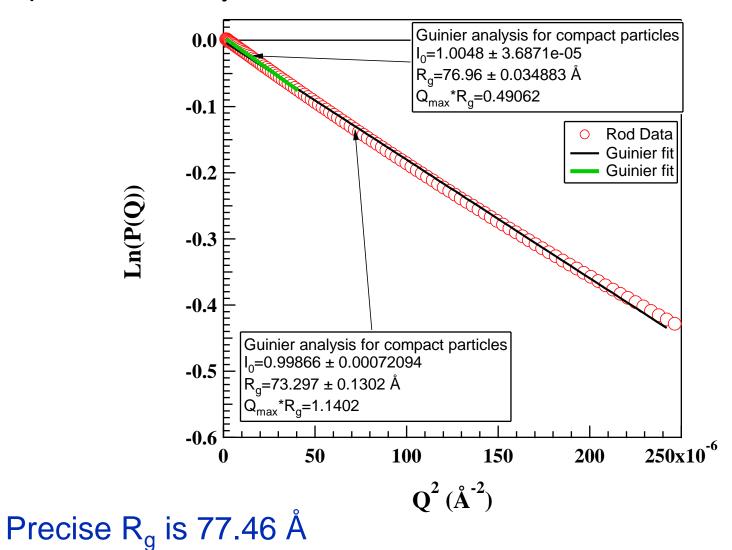
K.-H. Tang, L. Zhu, V. S. Urban, A. M. Collins, P. Biswas, and R. E. Blankenship, "Temperature and Ionic Strength Effects on the Chlorosome Light-Harvesting Antenna Complex," *Langmuir* 27 (8), 4816–4828(2011).



C. aurantiacus chlorosome (green) and model ellipsoid form (red) calculated from neutron scattering data of chlorosome entrapped in silica gel. Green volume reconstructed from electron density map of free chlorosome reported by Psencik et al.

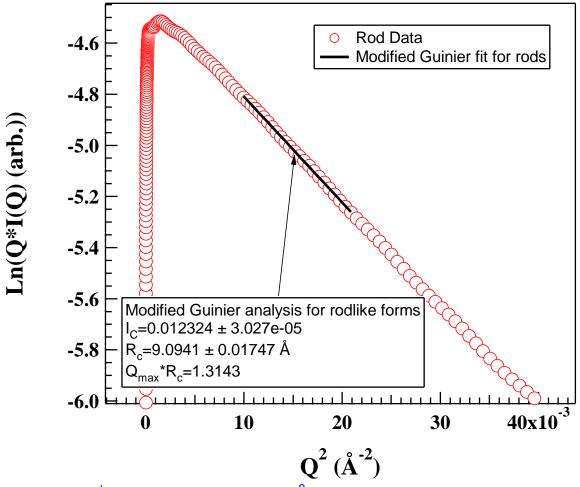
W.B. O'Dell, K.J. Beatty, J.K.H. Tang, R.E. Blankenship, V.S. Urban and H. O'Neill, "Sol-gel entrapped light harvesting antennas: immobilization and stabilization of chlorosomes for energy harvesting," *J. Mater. Chem.* 22(42), 22582-22591 (2012).

Guinier Analysis size of any kind of object





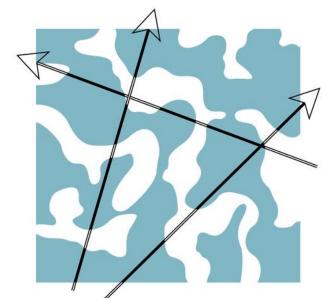
Modified Guinier Analysis for object extended in 1 dimension



Rod radius = $\sqrt{2}$ * R_c = 12.9 Å, exact radius = 13.3 Å A similar approach exists for thickness of (2d) sheet-like structure.

Non-particulate Scattering

Debye Bueche Model for Two-Phase System, Each with Random Shape, Uniform Electron or Scattering Length Density and Sharp Boundaries



Physical Concept of the Mean Chord or Inhomogeneity Length

Mean Chord Intercepts:

$$L_1 = \frac{a}{\phi}$$

$$L_2 = \frac{a}{(1-\phi)}$$

The fluctuations in scattering power at two points A and B, distance r apart, can be characterized by $\gamma(r) < \eta^2 >_{AV} = < \eta_A \eta_B >_{AV}$. For random two phase system: $\gamma(r) = e^{-r/a}$

$$\frac{d\Sigma}{d\Omega} (Q) = \frac{A}{[1 + Q^2 a^2]^2}$$

J. Appl.Cryst., 28, 679 (1957)

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